



Publishing Proteomics Data: How to Win Reviewers and Influence Editors

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St. Louis - ASMS 2015



San Antonio - ASMS 2016

Why do we publish?



https://computervisionblog.files.wordpress.com/2012/02/publication.jpg

Dissemination Education Promotion/tenure/funding Masochism?

Who do we need to satisfy?



https://psychanded.files.wordpress.com/2009/10/peer-review2.png



http://blog.historians.org/images/681.jpg

For submission, be prev Abstract Introduction Provide a list of "pr **STOP Nethods** Pick people w the area of / **ults and Discussion** Don't coun clusions on you. The **READ THE** borting information less than s Don't pick t **INSTRUCTIONS** They most review and FOR AUTHORS process. Provide a list of peo will be biased against

For submission, be prepared to:

Provide a list of "preferred reviewers."

Pick people who are knowledgeable in the area of the study.

Don't count on your friends being easy on you. They don't want you to publish a less than stellar paper.

Don't pick the "top names" in the field. They most likely won't have time for the review and it will delay the evaluation process.

Provide a list of people who you truly feel will be biased against your work.

Abstract Introduction Methods Results and Discussion Conclusions Supporting information

Succintly describe your project and results.

This may be all that the editor reads before inviting reviewers.

Abstract

Introduction

Methods

Results and Discussion

Conclusions

Supporting information

Here's your chance to make the case for your study. Keep on topic. This isn't a review article.



Abstract

Introduction
Methods
Results and Discussion
Conclusions
Supporting information

http://1.bp.blogspot.com/_9cOxMQ3Smgw/S53ZEN8FdPI/AAAAAAAAAXk/O16IPLI1EM8/s320/DV-revision.gif

Now the details . . .

How much is enough?

Abstract Introduction Methods Results and Discussion Conclusions Supporting information

Methods section

Protein analysis by mass spectrometry in the separated by 1-D SDS-PAGE and the gel lanes were dimensioned by the digests were and the digests were and the digest of the

Methods section

Rodriguez KA, Osmulski PA, Pierce A, Weintraub ST, Gaczynska M, Buffenstein R. A cytosolic protein factor from the naked mole-rat activates proteasomes of other species and protects these from inhibition. *Biochim Biophys Acta*. 2014 Nov;1842(11):2060-72. doi: 10.1016/j.bbadis.2014.07.005

	50 (1	50(1)		¢	analyzed by capital y in the electrospray ionization tandem mass spec-				
	NMR	MS	NMR	MS	trometry (HPLC-ESI-MS/MS) on a Thermo Fisher Orbitrap Velos mass				
Cytosolic/Adh	(VS) $125.0 \pm 5.6^*$	$5.56 \pm 1.57^{*}$	$80.6 \pm 2.5^{*}$	$16.2 \pm 1.9^{*}$	spectrometer. The MS data were searched against the rodent subset of				
Cytosolic/MG1	132 122.0 \pm 5.5 [*]	$8.22 \pm 2.75^{*}$	$71.4\pm8.7^{*}$	$14.6 \pm 4.4^{*}$	the NCBInr protein database (NCBInr_20130102; 316,972 sequences)				
Cytosolic/LC	0.1.1 D								
Cytosolic/B2 Microsomal	2.11. Prote	ein ident	ificatio	n by ma	iss spectrometry				
Nuclear/MG				-					
- = no data:									
(methyl)-sulf	Protei	ns were	senara	ated by	1-D SDS-PA				
LC = lactacys	TIOUCI		Separa	accu by	I D SDS IV				
* p > 0.05.	lane were	e digest	ed in s	<i>itu</i> with	n trypsi				
	1 1	1	1 11						
com/suppor	analyzed	by capil	lary H	PLC eleg	The ation tandem mass spec-				
1 ug of anti-	trometry (HDLC_ESL_MS/MS)								
or naked m	tronnetry	(III LC-		0/1VI	au l'islici Orbitrap velos mass				
After an	spectrom	eter. Th	۳ e MS		arched against the rodent subset of				
protein A/O	the NCPI	ar proto			(Plnr, 20120102; 216.072; coquences)				
bande ware	the NCDI	II prote			(CDIIII_20150102, 510,972 sequences)				
	by Masco	t		(e). T	he Mascot results were subjected to a				
	and hast as				llaurad has determined and a formal ability				
	subset se		r an	idem to	nowed by determination of probability				
	acceccme		ne nent	ide assi	gnments and protein identifications by				
	assessme	u u	ic pepi	iuc assi	ginnents and protein identifications by				
	Scaffold (Proteon	ne Soft	ware).					
	contoria (

Methods section - samples

Easy to provide

Source Animals Species Strain/genetic background Cells Cell line name designation Source Genetic alteration(s) Labeling strategy (SILAC, iTRAQ/TMT, SILAM) Number of biological and/or technical replicates

Methods section - sample preparation

Easy to provide

Protein isolation

Proteolytic digestion (if used)

Internal standards added (if used)

Chemical modification (if used), including labeling scheme

Off-line chromatography/cleanup

PTM-specific enrichment/isolation

Methods section - MS analysis

Easy to provide

HPLC

Instrument vendor/model

Column, mobile phases, flow rate, gradient, auxiliary detection details

Strategy for sample injection order

Mass spectrometry

Instrument vendor/model

Instrument parameters/scan strategy

HPLC-ESI-tandem-MS

Data-dependent analysis: MS1 mass resolution, MS1 scan range, charge-state screening parameters, mass window for precursor ion isolation, fragmentation mode, relative collision energy (or other parameter, as appropriate), mass analyzer for tandem-MS, MS2 mass resolution (where appropriate), number of product ion spectra per scan cycle, dynamic exclusion

Data-independent analysis: MS1 mass resolution, MS1 scan range(s), relative collision energy, mass analyzer for tandem-MS, MS2 mass resolution (where appropriate)

Methods section - data processing

Easy to provide

Software/method for peak list generation

Database searching

Software name(s), vendor(s) or literature citation, version

Databases

Name/source

Date/version

Taxonomy

Number of sequences

Search parameters

Precursor and product ion mass tolerances Enzyme specificity Charge states considered Fixed and variable modifications Other settings (*e.g.*, ¹³C number in Mascot) Quantitation method (where applicable) Decoy search option

Do not say "using the default parameters"

Methods section - data processing

Getting harder

Criteria for acceptance of peptide assignments and protein identifications

Mascot ions scores/expect values

SEQUEST X_{corr} cutoffs

X! Tandem scores

Post-processing with additional software

PeptideProphet/ProteinProphet

Scaffold

Proteome Discoverer

In-house software (need to document)

False discovery rate (FDR) determination

Decoy database details

Method for FDR calculation

De novo sequencing

Approach (*e.g.*, manual or computational)

Validation

Methods section - quantitative analysis

Getting even harder

Software name, vendor or literature citation, version Quantitation parameters Normalization

Methods section - quantitative analysis

Getting even harder

Considerations for acceptance of peptide values

SILAC/metabolic labeling

Variability of ratios across an EIC

Fraction of the EIC window

Agreement with predicted isotope pattern

Reporter ion-based methods

Variability assessment for reporter ion ratios of within-sample replicates

Spectral counting

Minimum number of assigned peptides/spectra

Spectrum designation

Total spectra, unique spectra, unique sequence

Intensity-based methods

Method for peak integration/intensity determination

Variability assessment for peptides assigned to a protein

Methods section - quantitative analysis

Getting even harder

Considerations for acceptance of peptide values

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Variability assessment for peptides assigned to a protein

Variability assessment of peptides assigned to a protein

		30 min		60 min			180 min			
tRNA snthetase	Accession	L/H	SD(geo)	#	L/H	SD(geo)	#	L/H	SD(geo)	#
Val	orf19.1295	3.13	2.60	89	26.93	4.81	62	7.92	1.19	47
lle	orf19.2138	4.13	3.77	33	28.75	3.92	16	7.07	1.35	22
Leu	orf19.2560	5.09	2.08	5	22.93	1.83	3	18.73	2.12	5
Ala	orf19.5746	5.94	1.09	6	15.36	13.60	3	5.76	1.11	4
Asn	orf19.6702	7.18	1.76	24	29.64	1.81	4	8.33	1.23	7
Gin	orf19.7064	5.55	1.51	11	4.95	31.09	17	5.91	17.21	16

Relative quantities of selected tRNA synthetases in *C. albicans* grown at different temperatures

Values were obtained from Mascot Distiller processing of the MS data and search results as multi-file projects for each time-point. H/L, median ratio of peak areas of extracted ion chromatograms for peptides assigned above the homology threshold; SD(geo), geometric standard deviation of the assigned peptides; #, number of peptides used for relative quantification.

Methods section - statistics

May be really difficult

Software/program Test(s) applied Significance levels



"I can prove it or disprove it! What do you want me to do?"

What did you find? How much documentation do you need to present - and where? How much documentation do you need to present - and where? How much documentation do you need to present - and where? How much documentation do you Supporting information

Abstract

Provide a clear, succint report of your results and insightful interpretations.

Highlight key findings. Do not cover every detail presented in the results tables and figures.

Focus on truly meaningful observations. Do you really need to discuss the biological signifcance of every protein you identified?

Results - tables of proteins

Table 3Mass spectrometry of fractions 22 and 23 revealed the presence of severalmolecular chaperones. For a complete list please see Table S1.

Identified proteins (8/223)	Accession number	MW	fr.22	fr.23
Inducible heat shock protein 70 (HSP72) [Heterocephalus glaber]	gi 13242237 (+26)	71 kDa	28	27
Ubiquitin-like modifier-activating enzyme 1 [Heterocephalus glaber]	gi 351699501	119 kDa	20	12
78 kDa glucose-regulated protein [Heterocephalus glaber]	gi 351702099	72 kDa	10	5
Inducible heat shock protein 70 [Mus musculus]	gi 118490060 (+7)	70 kDa	8	6
Hsp90aa1 protein [<i>Mus musculus</i>]	gi 118142832 (+23)	66 kDa	8	7
Heat shock protein 90 beta [Equus caballus]	gi 12082134 (+17)	82 kDa	7	8
Protein disulfide-isomerase [Heterocephalus glaber]	gi 351706419	57 kDa	7	5
Stress-70 protein, mitochondrial [Mus musculus]	gi 162461907 (+9)	73 kDa	7	6

Spectral counts (quantitative value) determined by Scaffold (v3) are shown in the table under the columns labeled fr.22 and fr.23.

Will readers know what this is?

What were the criteria for acceptance of peptide assignments and protein identifications?

Was an FDR assessed? How was it used?

Rodriguez, KA et al. Biochim Biophys Acta. 1842:2060-2072, 2014

Bacteriophage 201¢2-1 sturcture and morphology





Cryoelectron micrograph

1-D SDS PAGE of phage 201\u00e92-1 structural proteins



1-D SDS PAGE of phage 201\u00f62-1 structural proteins



76 proteins identified ≥2 peptides/protein Scaffold post-processing 95% confidence (peptides) ≥99.9% probability (protein)

Virology. 2008 Jul 5;376(2):330-8. doi: 10.1016/j.virol.2008.04.004



Characterization of *Pseudomonas chlororaphis* myovirus $201\phi^2-1$ via genomic sequencing, mass spectrometry, and electron microscopy

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A R T I C L E I N F O

Article history: Received 13 December 2007 Returned to author for revision

ABSTRACT

Pseudomonas chlororaphis phage 201 ϕ 2-1 is a relative of *Pseudomonas aeruginosa* myovirus ϕ KZ. Phage 201 ϕ 2-1 was examined by complete genomic sequencing (316,674 bp), by a comprehensive mass spectrometry survey of its virion proteins and by electron microscopy. Seventy-six proteins, of which at least 69

Table 1 MS data and homologues for the 201¢2-1 proteins identified by HPLC-ESI-MS/MS

			Identification by MS ¹			
gp	M _r (kDa)	Unique peptides	Total spectra	spectra Spectra/Mw % coverage		Function, homologues, paralogues ²
A. Proteins of	f established virion f	function according to Fokin	ne et al. (2007)			
30 ³	77.5	22	355	4.6	44	major sheath protein, KZ ⁴ 29 (63% over 693); EL6 (21% over 707)
200 ⁵	82.4	22	696	8.4	38	major capsid protein, KZ120 (64% over 749); EL78 (20%
276 ⁵	251.8	23	34	0.1	10	cell-puncturing device, KZ181 (33% over 2387); KZ144 (45% over 187); EL183 (22% over 270)
B. RNA polyn	nerase-related virion	n proteins				
139	49.6	8	34	0.7	24	RNA polymerase, beta' subunit, KZ80 (56% over 449); EL44
273/274	173.4	24	87	0.5	19	RNA polymerase, beta subunit, KZ178 (53% over 1548);
275	62.7	8	27	0.4	15	EL180 (25% over 1142); EL187 (26% over 556) RNA polymerase, beta subunit KZ180 (68% over 490); EL184 (32% over 491)

¹All proteins had a protein identity probability of 100%, as determined by Scaffold (Proteome Software), with the exception of gp164 (99%) and gp229 (96%). Results displayed were obtained from a combined data set of the GeLCMS analysis, with the exception of gp276 which was only detected in analysis of an individual gel band (see text).

²Homologues were determined using Psi-Blast and BlastP (% identities over the homologous region are provided). The best matching φKZ and EL homologue for each 201φ2-1 protein is listed. Paralogue families are as follows: paralogue family a refers to a domain found in 201φ2-1 gp216, 217, 218, 219, 220. A homologous domain exists in φKZ gp131, 132, 133, 134 and 135 and EL gp113, 114 and 115; paralogue family b refers to a domain found in 201φ2-1 gp155, 156, 157, 246, 247. Homologous domains exist in φKZ gp93, 94, 95, 162 and 163; paralogue family c refers to a domain found in 201φ2-1 gp155.

³An N-terminal peptide lacking only the initiator methionine was identified using semi-tryptic analysis.

⁴KZ refers to φKZ.

 5 N-terminus is expected to be processed as the ϕ KZ homologue is processed. Although no semi-tryptic fragments were found to define the mature ends, there is also a lack of peptide coverage in the N-terminal region that would be consistent with processing.

 $^{\it O}$ A mature N-terminus containing the initiator methionine was confirmed by semi-tryptic analysis.

⁷Gel analysis indicated that the protein is processed consistent with a lack of MS sequence coverage in the N-terminal region of these sequences, except for gp246N which lacks MS coverage of the C terminal region. The exact positions of the processed ends are unknown. The normalized spectrum count in parentheses was calculated using the apparent molecular weight of the processed form (Fig. 1).

⁸Semi-tryptic analysis indicated removal of 63 and 60 N-terminal residues from gp238 and gp271, respectively.

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273/274	173.4	24	87	0.5	19	(25% over 447) RNA polymerase, beta subunit, KZ178 (53% over 1548);
275	62.7	8	27	0.4	15	EL186 (25% over 1142), EL187 (26% over 556) RNA polymerase, beta subunit KZ180 (68% over 490); EL184 (32% over 491)

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⁸Semi-tryptic analysis indicated removal of 63 and 60 N-terminal residues from gp238 and gp271, respectively.



Think about what you're including as supporting data.

Make sure the figures and tables are informative.

Use meaningful titles for the figures and tables.

Don't provide every spreadsheet you or the software generated as a part of the analysis.

Rearrange spreadsheet columns as needed so that the key information is easily found.

What needs to be documented?

There is no concensus about whether all identified proteins need to be listed or only those that exhibited significant differences in quantity among experimental groups or are of special interest to the study.

Identification by HPLC-ESI-tandem-MS

Protein level report - spreadsheet format, PLEASE

Columns

Protein name

Accession number

Molecular weight

Number of assigned spectra (total and unique)

Percent sequence coverage

Probability of protein inference (if determined during post-processing)

Clear/meaningful column heading names

Legend - at the top of long tables or on a separate worksheet Explanations for abbreviated or non-standard column headings Significance level cutoff of assigned peptides

MALDI-TOF-MS (peptide mass fingerprint, PMF)

- Only suitable for low-complexity samples
- Protein level report
 - Columns
 - **Protein name**
 - Accession number
 - Molecular weight
 - Number of spectra searched
 - Number of spectra assigned
 - Percent sequence coverage
 - **Probability of protein inference**
 - **Clear/meaningful column heading names**
 - Legend at the top of long tables or on a separate worksheet Explanations for abbreviated or non-standard column headings

MALDI-TOF/TOF

Protein level report Columns **Protein name** Accession number Molecular weight Number of spectra searched for PMF Number of spectra assigned for PMF Number of tandem-mass spectra searched Number of tandem-mass spectra assigned Percent sequence coverage **Probability of protein inference Clear/meaningful column heading names** Legend - at the top of long tables or on a separate worksheet Explanations for abbreviated or non-standard column headings Significance level cutoff of assigned peptides

Peptide level (grouped by protein)

For documentation of PTMs, sequence variations

Not usually necessary for manuscripts focusing on identification or relative quantification

Columns

Protein name

Peptide sequence - clearly showing modification(s)

Start/stop residue numbers

Observed *m/z*

Charge state

Mass error

Score/expect value for sequence assignment

Probability for localization of modification site (where appropriate) Clear/meaningful column heading names

Legend - at the top of long tables or on a separate worksheet Explanations for abbreviated or non-standard column headings Significance level cutoff of assigned peptides

Supporting information Annotated tandem mass spectra

There is no concensus about when annotated tandem mass spectra need to be provided.

Post-translational modifications

Do we need to see all spectra for a large phosphoproteomics study if acceptable parameters have been used for database searching and reasonable cutoffs have been applied to site localization probabilities? Unusual modifications or surprising findings that are the focus of the manuscript should be documented in the body of the manuscript.

Proteins identified by a single, high-confidence peptide assignment Will examining the annotated spectra influence confidence about the assignment? Will you really look at them?

Annotated tandem mass spectra - annotate the following for each

Peptide sequence Observed *m/z* Mass error Charge state Database search score Probability/expect score Site localization probability (where appropriate)
Supporting information Annotated tandem mass spectra

Protein family 139 transketolase [Trichoderma atroviride IMI 206040] Score, 106; matches, 2; match(sig), 2; sequences, 2; seq(sig), 2; eMPAI, 0.06



Print a PDF of the Mascot Peptide View

Peptide View

MS/MS Fragmentation of LEGILPELVGGSADLTGSNITP

Found in gi|358391264 in NCBInr, transketolase [Trichot All matches to this query

Match to Query 6007: 2211.175448 from(1106.595000,2+ Title: 5776: Scan 10850 (rt=34 9916) [\\lcabio\MSL 1\Wo

	Score	Mr(calc)	Delta	Sequence					
20 1 4 1 4 1 4 1 4 4 4 4 4 4 4 4 4 4 4 4	86.6	2211.1747	0.0008	IEGILPELVGGSADLTGSNLTR					
LEGILPELVGGSADLTG	86.6	2211.1747	0.0008	LEGILPELVGGSADLTGSNLTR					
	16 .5	2211.1648	0.0107	<u>GLITVTYDVNPRQIDIHTR</u>					
	15.1	2211.1648	0.0107	<u>GLITVTYDVNPRQIDIHTR</u>					
eak **(8)	11.0	2211.1787	-0.0032	IDAILKAGFADLHVITEEQK					
p (c c c c c c c c c c c c c c c c c c c	9.7	2210.1439	1.0316	MNAQVLNLVAALGVMQYSKK					
% of l % of l (6) ((6) (18) ⁺⁺ (18) ⁺⁺	9.2	2211.1821	-0.0066	LLGELLGAVTDPVCNLLTGQK					
20 - (+)q (5)	9.1	2211.1470	0.0284	LGSHMPLEDFLQLLRDATR					
	8.9	2211.1324	0.0430	LAINIWYKPAGGSPPTPDGTR					
expect value matches	8.8	2211.1510	0.0244	VSLLIPPWICHSFNAISTR					
(2) [40.18]									



Supporting information Annotated tandem mass spectra

Protein family 376 uroporphyrinogen decarboxylase [Cryptococcus neoformans var. grubii H99] Score, 30; matches, 1; match(sig), 1; sequences, 1; seq(sig), 1; eMPAI, 0.07





Peptide View

MS/MS Fragmentation of VHSVLSQLSHPGVPITLFAK

Found in gi|405118227 in NCBInr, uroporphyrinogen dec

ec All matches to this query

Match to Query 5835: 2129.199492 from(710.740440,3+) Title: 5615: Scan 10667 (rt=34.4181) [\\lcqbio\MSL 1\Wo



🔄 🔍 🔍 🔍 🔣 120.12 to 1368.74 🔍 🤅

Monoisotopic mass of neutral peptide Mr(calc): 2129.1997 Fixed modifications: Carbamidomethyl (C) (apply to specified residues or termini only) Ions Score: 30 Expect: 0.023 Matches : 26/204 fragment ions using 33 most intense peaks (help)

Supporting information

Quantitative analysis

Protein level report

Documentation can be added to the ID report or in a separate table

Columns

Protein name

Accession number

Molecular weight

Number of assigned spectra (total and unique)

Percent sequence coverage

Probability of protein inference (if determined post-processing)

Number of peptides used for quantitative analysis

Variability of results for peptides assigned to a protein

Clear/meaningful column heading names

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Legend - at the top of long tables or on a separate worksheet Explanations for abbreviated or non-standard column headings Significance level cutoff of assigned peptides

Supporting information

You can generate the supplemental data tables needed for publication directly from Mascot by exporting a CSV file from the Mascot Results Report page.

$\Delta cap \Delta cap D$ 265 D MW H99 H99 D 265 kDa 6 140 115 5 80 65 4 50 40 3 30 2 25 1 15 В С D Α Ε F BioRad Criterion XT MOPS 12% SDS-PAGE reducing gel, blue silver stain

1-D SDS PAGE - proteins of *Cryptococcus neoformans*

A.K. Chaturvedi and F.L. Wormley, Jr.

Mascot search engine Protei × Wormley Chaturvedi M1415-058 1 × +			↔ _	. 8 ×
🔹 🕲 vprmascot. win. uthscsa.edu/mascot/cgi/master_results_2.pl?file=.,%2Fdata%2F20150515%2FF035682.dat;sessionID=weintraub_264016471540778 🔍 C	a 🔸	俞	ø	≡
🧕 Most Visited 🗌 Getting Started 🎆 Weather Forecast Sa 😑 Examsoft Dental 🦳 Austin-San Antonio R 🗌 Patient Portal 🧿 Intellicast - San Anto 🦹 The University of Tex 🗽 Mascot_vprmascot				
User : stweintraub E-mail : weintraub@uthscsa.edu Search title : Wormley Chaturvedi M1415-058 150130FLW32_F3.raw (NCBInr_fungi; contam) trypsin-1 Ox(M) Carb(C) D(NQ) decoy (Wormley Chaturvedi M1415-058 265 D, slice F3, 5ul, D Databases : 1: contaminants 20120713 (247 sequences; 128,130 residues) 2: NCBInr 20140522 (39,649,990 sequences; 14,178,194,136 residues))8)			
Taxonomy : 1: (none) 2: Fungi (2,653,696 sequences) Timestamp : 15 May 2015 at 19:07:55 GMT				
Re-search • All C Non-significant C Unassigned @[help] Export As XML				
Not what you expected? Try zithe select summary.				
Search parameters				
Score distribution				
Modification statistics				
r Legena				
Protein Family Summary				
Filter Significance threshold p< 0.05 Max. number of families AUTO Implement Ions score or expect cut-off 0 Dendrograms cut at 0 0 Show Percolator scores Implement Allentries Implement Implement Preferred taxonomy Allentries Implement Implement Implement				
Decoy search summary (reversed protein sequences)				
Proteins (389) Report Builder Unassigned (5102)	{	§ perr	malin	<u>ık</u>
Protein families 1–10 (out of 389)				
10 v per page 1 2 3 4 5 6 39 Next Expand all Collapse all				
1 2::gi 58261082 1222 14-3-3 protein [Cryptococcus neoformans var. neoformans JEC21] 2 3::gi 521772019 500 hupathetical archein BCT05224 020 [Blumaria araminia f an kritici 05224]				•

Mascot search engine Protei × Wormley Chaturvedi M1415-058 1 ×	+				-	•	8 >
(vprmascot.win.uthscsa.edu/mascot/cgi/master_results_2.pl?file=%2Fdata%	2F20150515%2FF035682.dat;sessionID=weintraub_264016471540778	⊽ C Q Search	☆ 自	÷	A	ø	≡
🔊 Most Visited 📋 Getting Started 🔜 Weather Forecast Sa 📄 Examsoft Dental	📄 Austin-San Antonio R 📄 Patient Portal 🧕 Intellicast - San Anto 🦹 The Univers	sity of Tex 🗽 Mascot_vprmascot					18
	esults						1
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scores > 48 (beyond green shading) indicate extensive homology (p<0.05).

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1	1	NCBInr	Accession 2010/01/58261082	1222	20000	112	61	27	18	64 20	14-3-3 protein [Cryptococcus peoformans var. peoformans]EC21]	—
1	2	NCBInr	2::gi[62420901	629	28911	72	33	17	10	12.65	14-3-3 1 protein [Phanerochaete chrysosporium]	
1	3	NCBInr	Z::gi 521773918	500	30146	50	29	10	8	9.60	hypothetical protein BGT96224 929 [Blumeria graminis f. sp. tritici 96224]	1
1	4	NCBInr	d'2::gi 254567754	161	29122	36	17	10	6	4.64	hypothetical protein [Komagataella pastoris GS115]	
1	5	NCBInr	data di alta di a	132	30471	22	9	7	5	2.00	rad24 [Schizosaccharomyces pombe]	
1	6	NCBInr	del::gi 50546823	61	31318	12	3	4	3	0.49	YALI0B14377p [Yarrowia lipolytica]	
2	1	NCBInr	delate delta della dell	688	85793	45	28	31	20	1.96	heat shock protein [Cryptococcus gattii WM276]	
2	2	NCBInr	ď 2::gi 405123671	488	85923	36	20	24	14	1.09	heat shock 70kDa protein 4 [Cryptococcus neoformans var. grubii H99]	
<u>2</u>	3	NCBInr	delate delta della dell	178	85983	8	5	5	4	0.22	hypothetical protein SERLADRAFT_355892 [Serpula lacrymans var. lacrym	ans
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Mascot search results report - ions score cutoff 0, no FDR adjustment

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29		1	NCBInr	2::ail321258651	378	35869	23	15	13	9	3.08	inorganic diphosphatase [Cryptococcus gattii WM276]
29		2	NCBInr	d2::qi 540383854	248	35443	20	14	11	9	2.69	inorganic pyrophosphatase [Cryptococcus neoformans var. grubii H99]
30		1	NCBInr	d2::gi 321260106	370	58177	32	19	20	13	1.76	ATP synthase alpha chain, mitochondrial precursor [Cryptococcus gattii WM27
31		1	NCBInr	d2::gi 321257325	365	39278	27	16	17	11	3.02	hypothetical protein CGB_D4230W [Cryptococcus gattii WM276]
31		2	NCBInr	⊿2::gi 134111162	294	39332	14	10	8	6	1.35	hypothetical protein CNBD4520 [Cryptococcus neoformans var. neoformans B-
32		1	NCBInr	⊿2::gi 321253911	363	34764	21	12	10	7	1.97	hypothetical protein CGB_C5340C [Cryptococcus gattii WM276]
<u>32</u>		2	NCBInr	₫2::gi 405119194	58	34698	9	3	6	3	0.44	aldose reductase [Cryptococcus neoformans var. grubii H99]
<u>33</u>		1	NCBInr	id 2::gi 321259223	349	48772	34	20	11	9	2.08	cellulase [Cryptococcus gattii WM276]
34		1	NCBInr	d 2::gi 321259373	319	39890	36	16	16	11	2.55	6-phosphogluconolactonase [Cryptococcus gattii WM276]
<u>34</u>		2	NCBInr	d2::gi 405120916	301	34650	25	13	13	9	2.36	6-phosphogluconolactonase [Cryptococcus neoformans var. grubii H99]
<u>35</u>		1	NCBInr	del::gi 321253449	303	69578	36	16	25	14	1.64	heat shock protein 70 [Cryptococcus gattii WM276]
35		2	NCBInr	id 2::gi 58264706	260	60520	25	15	24	12	1 24	beat check protein 70 [Countercore]us neoformans var. neoformans JEC21]
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35		4	NCBInr	ď2::gi 121568		10101	ii iai					gulated protein homolog; Short=GRP-78; Al
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35		8	NCBInr	Z::gi[599407270			S	core		256		_265825 [Phanerochaete carnosa HHB-1011
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40		1	NCBInr	2::gi 321257249			S	equer	nces	13		rotein [Cryptococcus gattii WM276]
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42		1	NCBInr	2:: gi 321253782	1		S	ea(sid	(r	7		[Cryptococcus gattii WM276]
43		1	NCBInr	d2::ai 321261545]		Ŭ	94(9)	5/	•		[Cryptococcus gattii WM276]
43		2	NCBInr	⊿2:: gi 134113809			e	MPAL	ſ) 87		ryptococcus neoformans var. neoformans B
44		1	NCBInr	₫2::gi 58258849			Ŭ		Ŭ			occus neoformans var. neoformans JEC21]
<u>45</u>		1	NCBInr	ď 2::gi 302907708	219	28241	16	7	6	4	1.44	predicted protein [Nectria haematococca mpVI 77-13-4]
<u>45</u>		2	NCBInr	ď 2∷gi 615413004	189	25535	10	5	5	3	0.93	putative deoxyribose-phosphate aldolase protein [Neofusicoccum parvum UCR
<u>46</u>		1	NCBInr	d 2::gi 321264171	219	31471	14	9	10	7	1.90	hypothetical protein CGB_K3180C [Cryptococcus gattii WM276]
<u>47</u>		1	NCBInr	⊿2::gi 321254861	218	52235	23	11	16	8	1.07	aminotransferase [Cryptococcus gattii WM276]
<u>47</u>		2	NCBInr	⊿2::gi 405119067	193	52462	16	9	13	7	0.90	4-aminobutyrate transaminase [Cryptococcus neoformans var. grubii H99]
10		1	MCRInr	20000158258141	217	26155	20	Q	0	5	1.61	hunothatical protain CNA00560 [Chuntacoccus poofarmans var. poofarmans IF*

Mascot search results report - ions score cutoff 0, no FDR adjustment



Unformatted sequence string: <u>488 residues</u> (for pasting into other applications).

Sort peptides by

Residue Number

Increasing Mass
Decreasing Mass

Show predicted peptides also

Query	Start - End	Observed	Mr (expt)	Mr(calc)	ppm M	Score	Expect	Rank	U	Peptide
₫5902	9 - 26	721.3964	2161.1673	2161.1671	0.13 1	37	0.0068	1	U	K.QIEPYFVLNDEKLVDIVK.H
₫ 7636	42 - 73	1123.5515	3367.6327	3367.6316	0.31 0	39	0.0079	1	U	K.DMAMIPTFVTGVPDGTEEGVFLALDLGGTNLR.V + 2 Oxidation (M)
₫7637	42 - 73	1123.8851	3368.6335	3367.6316	297 0	41	0.0077	1	U	K.DMAMIPTFVTGVPDGTEEGVFLALDLGGTNLR.V + 2 Oxidation (M)
₫4184	74 - 86	766.9161	1531.8175	1531.8181	-0.35 0	66	0.00093	1	U	R.VCLIVLQGNNQFK.I
₫4185	74 - 86	511.6135	1531.8186	1531.8181	0.36 0	40	0.46	1	U	R.VCLIVLQGNNQFK.I
1885	176 - 183	489.2554	976.4963	976.4978	-1.54 0	41	0.15	1	U	R.LLQDAFDR.K
₫2551	176 - 184	369.2042	1104.5908	1104.5927	-1 76 1	27	0.98	1	U	R.LLQDAFDRK.H
₫4467	190 - 204	803.4171	1604.8195	1604.8192	0.21 0	100	1.8e-007	1	U	R.CSALVNDTVGTLLSR.S
₫4468	190 - 204	535.9474	1604.8204	1604.8192	0.72 0	45	0.12	1	U	R.CSALVNDTVGTLLSR.S
₫6396	205 - 228	801.4020	2401.1841	2401,1801	1.64 0	59	0.008	1	U	R.SYOSGPALIGAIFGTGTNGAYIDK.S + Deamidated (NO)

Score	Expect Rank	U	Peptide
37	0.0068 1	U	K.QIEPYFVLNDEKLVDIVK.H
39	0.0079 1	U	K.DMAMIPTFVTGVPDGTEEGVFLALDLGGTNLR.V + 2 Oxidation (M)
41	0.0077 1	U	K.DMAMIPTFVTGVPDGTEEGVFLALDLGGTNLR.V + 2 Oxidation (M)
66	0.00093 1	U	R.VCLIVLQGNNQFK.I
40	0.46 1	U	R.VCLIVLQGNNQFK.I
41	0.15 1	U	R.LLQDAFDR.K
27	0.98 1	U	R.LLQDAFDRK.H
100	1 8e-007 1	п	R CSALVNDTVGTLLSR S

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20	1			EL.Ighteleosoer	501	55150		10			1.20	
29	1	NCBInr		■2::gi 321258651	378	35869	23	15	13	9	3.08	inorganic diphosphatase [Cryptococcus gattii WM276]
29	2	NCBInr	1	≥2::gi 540383854	248	35443	20	14	11	9	2.69	inorganic pyrophosphatase [Cryptococcus neoformans var. grubii H99]
30	1	NCBInr	4	≥2::gi 321260106	370	581//	32	19	20	13	1.76	ATP synthase alpha chain, mitochondrial precursor [Cryptococcus gattii WM27
<u>31</u>	1	NCBInr	7	d2::gi 321257325	365	39278	27	16	17	11	3.02	hypothetical protein CGB_D4230W [Cryptococcus gattii WM276]
<u>31</u>	2	NCBInr	2	⊠2::gi 134111162	294	39332	14	10	8	6	1.35	hypothetical protein CNBD4520 [Cryptococcus neoformans var. neoformans B
<u>32</u>	1	NCBInr	100	2::gi 321253911	363	34764	21	12	10	7	1.97	hypothetical protein CGB_C5340C [Cryptococcus gattii WM276]
32	2	NCBInr	2	≥2::gi 405119194	58	34698	9	3	6	3	0.44	aldose reductase [Cryptococcus neoformans var. grubii H99]
<u>33</u>	1	NCBInr	1	data array and a second secon	349	48772	34	20	11	9	2.08	cellulase [Cryptococcus gattii WM276]
<u>34</u>	1	NCBInr		delate all sectors and sector	319	39890	36	16	16	11	2.55	6-phosphogluconolactonase [Cryptococcus gattii WM276] —
<u>34</u>	2	NCBInr	- 2	≥ 2::gi 405120916	301	34650	25	13	13	9	2.36	6-phosphogluconolactonase [Cryptococcus neoformans var. grubii H99]
<u>35</u>	1	NCBInr	1	del::gi 321253449	303	69578	36	16	25	14	1.64	heat shock protein 70 [Cryptococcus gattii WM276]
35	2	NCBInr	1	ď2∷gi 58264706	269	69538	35	15	24	13	1.34	heat shock protein 70 [Cryptococcus neoformans var. neoformans JEC21]
35	3	NCBInr	8	data ang ang ang ang ang ang ang ang ang an	231	71915	11	6	10	5	0.34	heat shock protein [Cryptococcus gattii WM276]
35	4	NCBInr	3	ď2::gi 121568	91	74459	4	3	3	2	0.12	RecName: Full=78 kDa glucose-regulated protein homolog; Short=GRP-78; Al
35	5	NCBInr	1.54	d 2::gi 321265704	85	82556	6	4	6	4	0.23	kar2 karyogamy protein [Cryptococcus gattii WM276]
35	6	NCBInr	2	2::gi 255712457	<mark>69</mark>	70557	6	2	4	2	0.13	KLTH0C06556p [Lachancea thermotolerans]
<u>35</u>	7	NCBInr		⊠'2::gi 134106591	64	72135	6	5	4	4	0.26	hypothetical protein CNBA3060 [Cryptococcus neoformans var. neoformans B-
<u>35</u>	8	NCBInr		ď 2∷gi 599407270	56	67308	6	5	4	4	0.29	hypothetical protein PHACADRAFT_265825 [Phanerochaete carnosa HHB-1011
<u>36</u>	1	NCBInr		2::ai 321260961	301	34696	22	11	12	7	1.63	L-malate dehydrogenase [Cryptococcus gattii WM276]
<u>37</u>	1	NCBInr		₫2::gi 259120714	256	54004	23	13	13	7	0.87	hexokinase 2 [Cryptococcus gattii]
<u>38</u>	1	NCBInr		<mark>≥2gi 321254</mark> 947	254	80372	48	18	28	15	1.32	catalase A [Cryptococcus gattii WM276]
<u>38</u>	2	NCBInr		ď 2∷gi 109156571	131	80456	34	10	16	7	0.44	catalase 1 [Cryptococcus neoformans]
<u>39</u>	1	NCBInr		ď 2∷gi 321265355	244	26827	21	15	7	6	3.07	hypothetical protein CGB_M3280C [Cryptococcus gattii WM276]
<u>39</u>	2	NCBInr		ď2∷gi 405123684	138	26927	17	9	5	4	1.17	allergen [Cryptococcus neoformans var. grubii H99]
<u>40</u>	1	NCBInr		ď2::gi 321257249	235	33267	23	12	13	7	2.53	fatty acid beta-oxidation-related protein [Cryptococcus gattii WM276]
<u>41</u>	1	NCBInr		ď2∷gi 321252988	230	38587	20	8	13	5	0.92	zinc-binding dehydrogenase [Cryptococcus gattii WM276]
<u>42</u>	1	NCBInr		ď 2∷gi 321253782	229	64153	13	8	9	5	0.69	hypothetical protein CGB_C5160W [Cryptococcus gattii WM276]
<u>43</u>	1	NCBInr		ď2∷gi 321261545	228	37514	10	7	6	5	1.19	hypothetical protein CGB_G1660W [Cryptococcus gattii WM276]
<u>43</u>	2	NCBInr		≥2::gi 134113809	212	36778	9	6	6	5	0.99	hypothetical protein CNBG1350 [Cryptococcus neoformans var. neoformans B-
<u>44</u>	1	NCBInr		ď 2∷gi 58258849	226	40775	12	7	7	3	0.51	isocitrate dehydrogenase [Cryptococcus neoformans var. neoformans JEC21]
<u>45</u>	1	NCBInr		ď 2∷gi 302907708	219	28241	16	7	6	4	1.44	predicted protein [Nectria haematococca mpVI 77-13-4]
<u>45</u>	2	NCBInr		data di anticia di an	189	25535	10	5	5	3	0.93	putative deoxyribose-phosphate aldolase protein [Neofusicoccum parvum UCR
46	1	NCBInr		data 2::gi 321264171	219	31471	14	9	10	7	1.90	hypothetical protein CGB_K3180C [Cryptococcus gattii WM276]
47	1	NCBInr		data di anticia di an	218	52235	23	11	16	8	1.07	aminotransferase [Cryptococcus gattii WM276]
<u>47</u>	2	NCBInr		ď2∷gi 405119067	193	52462	16	9	13	7	0.90	4-aminobutyrate transaminase [Cryptococcus neoformans var. grubii H99]
10	1	NCRIDE			217	26155	20	Q	n	5	1 61	hypothetical protein CNA00560 [Chyptococcus peoformans var. peoformans]F

Mascot search results report - ions score cutoff 0.05, no FDR adjustment



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+ @ vprmascot.win.uthscsa.edu/mascot/cgi/master	_results_2.pl?file=%2Fdata%2F20150515%2FF035682.dat;_ignoreionsscorebelow=0;_prefertaxonomy=0;_sigthreshold=0.05;percolate=0;pr.show=reportbuilder; 🛡 🖸 🔍 Search	2 自 🕹 🏠 😕 🚍									
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E-mail : weintraub@uthscsa.edu Search title : Wormley Chaturvedi M1415-058 150130FLW32_F3.raw (NCBInr_fungi; contam) trypsin-1 0x(M) Carb(C) D(NQ) decoy (Wormley Chaturvedi M1415-058 265 D, slice F3, 5ul, D8) Datases : 1: contaminants 20120713 (247 sequences; 128,130 residues) 2: NCBIrr 201407527 (39 649 990 sequences; 14 178 194 136 residues)											
2: NCBIII 20140522 (39,649,990 Sequences; 14,178,194,136 residues) Taxonomy : 1: (none) 2: Fungi (2,653,696 sequences) Timestamp : 15 May 2015 at 19:07:55 GMT											
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Adjust FDR to 1% (or other desired value)



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Influence of Mascot significance and FDR settings

Hexokinase 2 [Cryptococcus gattii] gi|259120714

Significance	lons score				Significant		Significant
Threshold	cutoff	FDR (%)	Score	Matches	Matches	Sequences	Sequences
0.05	0	(4.2)	256	23	13	13	7
0.05	0.05	(4.2)	256	13	13	7	7
0.0122	0	0.94	194	23	9	13	6
0.0122	0.05	0.94	194	13	9	7	6







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	r <mark> ⊭2::gi 1040756 92 30471 8 6</mark>	4 3		rad24 [Schizosaccharomyces pombe]

Export as CSV

Mascot results spreadsheet



Give worksheets informative names.

Database column can be deleted if only one database was used.

Differential expression analysis using SILAC (Stable Isotope Labeling with Amino Acids in Cell Culture)



Assessment of differences in protein expression of human cells infected with Marburg viruses using SILAC

Lethality: Marburg virus-Angola (MARV-Ang) > Marburg virus-Musoke (MARV-Mus)



BioRad Criterion XT MOPS 12% SDS-PAGE reducing gel, "blue silver" stain

Mascot Distiller results



Ready

Quantitation table (proteins)

	L/H	SD(geo)	#	Description
1.1	0.8897	1.1602	136	fatty acid synthase [Homo sapiens]
1.2	0.8574	1.0880	19	PREDICTED: similar to fatty acid synthase [Canis familiaris]
1.3	0.8831	1.0685	2	fatty acid synthase [Ovis aries]
2.1	1.0599	1.0757	44	PREDICTED: glyceraldehyde-3-phosphate dehydrogenase [Callithrix jacchus]
2.2	1.1083	1.5353	31	glyceraldehyde-3-phosphate dehydrogenase [Phoca largha]
2.3	1.1096	1.5338	31	PREDICTED: similar to glyceraldehyde-3-phosphate dehydrogenase [Monodelphi
2.4	1.0233	1.1004	22	RecName: Full=Glyceraldehyde-3-phosphate dehydrogenase; Short=GAPDH; Al
2.5	1.1107	1.0731	22	PREDICTED: similar to glyceraldehyde-3-phosphate dehydrogenase [Canis famili
2.6	1.1118	1.0799	17	glyceraldehyde-3-phosphate dehydrogenase [Oryctolagus cuniculus]
2.7	1.0156	1.0956	15	Glyceraldehyde-3-phosphate dehydrogenase [Mus musculus]
2.8	1.0150	1.1043	14	PREDICTED: similar to glyceraldehyde-3-phosphate dehydrogenase [Monodelphi
2.9	1.1120	1.0702	7	PREDICTED: similar to Glyceraldehyde-3-phosphate dehydrogenase (GAPDH) [
2.10	1.1245	1.9014	13	PREDICTED: similar to glyceraldehyde-3-phosphate dehydrogenase [Canis famili
2.11	0.9968	1.0179	6	Chain R, Twinning In Crystals Of Human Skeletal Muscle D- Glyceraldehyde-3-P
2.12	0.9780	1.0256	5	PREDICTED: glyceraldehyde-3-phosphate dehydrogenase-like [Callithrix jacchus]
3	1.2503	1.0930	81	spectrin alpha chain, brain isoform 3 [Homo sapiens]
4.1	1.2820	1.1059	70	beta-spectrin [Homo sapiens]
4.2	1.2789	1.1109	66	spectrin beta chain, brain 1 isoform 2 [Homo sapiens]
4.3	1.2936	1.0071	3	spectrin-like protein GTRAP41 [Rattus norvegicus]
5.1	1.8338	1.0796	63	myosin-9 [Homo sapiens]
5.2	1.8338	1.0803	51	PREDICTED: LOW QUALITY PROTEIN: myosin-9 [Sus scrofa]
5.3	1.8201	1.0813	50	myosin, heavy polypeptide 9, non-muscle [Rattus norvegicus]
5.4	0.9973	1.0774	23	PREDICTED: myosin, heavy polypeptide 10, non-muscle isoform 8 [Pan troglodyt
5.5	1.8478	1.0829	4	PREDICTED: myosin, heavy polypeptide 14 isoform 4 [Pan troglodytes]


Ready

Peptide matches: fatty acid synthase (protein 1.1)

		z	Sequence	X	L/H	Std.Err	Fraction	Correlation	Intensity	Modificatio
1	+	2	LMSAISK	Г	1.1126	0.18201	0.4552	0.9847	98570	Oxidation (M)
2	+	2	TPEAVQK		0.9070	0.00982	0.7756	0.9957	738628	
3	+	2	DIMLATGK		0.8905	0.02451	0.7680	0.9902	365249	
4	÷	2	RAYLQAR	Г	0.7554	0.07658	0.4302	0.9802	156972	
5	+	2	GYAVLGGER	Г	3.6706	0.27966	0.4606	0.8926	30388	
6	+	2	KLQELSSK		0.7947	0.01824	0.6528	0.9920	783772	
7	+	2	HGLYLPTR		0.8005	0.01287	0.5122	0.9961	1663546	
8	+	2	HGLYLPTR	Г	0.8910	0.04431	0.4473	0.9946	343135	
9	÷	2	GLVQALQTK		0.8556	0.01614	0.9430	0.9976	1585877	
10	+	2	GLVQALQTK	Г	1.4824	0.06835	0.4623	0.9925	25356	
11	+	2	TGTVSLEVR	Г	0.8174	0.02263	0.3872	0.9970	232042	
12	+	2	TGTVSLEVR	Г	1.1870	0.05471	0.2344	0.9793	27681	
13	+	2	VLEALLPLK		0.9445	0.02065	0.8568	0.9962	432986	
14	÷	2	VLEALLPLK		0.8698	0.00885	0.9534	0.9986	2234177	
		n.	S ALE ALL ENTRY		4 4 C C C C	0.01000	0.0710	0.0000	00000	

Peptide matches: fatty acid synthase (protein 1.1)

		z	Sequence	X	L/H	Std.Err	Fraction	Correlation	Intensity	Modificatio
4	+	2	RAYLQAR	Г	0.7554	0.07658	0.4302	0.9802	156972	
5	÷	2	GYAVLGGER	Г	3.6706	0.27966	0.4606	0.8926	30388	
6	÷	2	KLQELSSK		0.7947	0.01824	0.6528	0.9920	783772	
7	÷	2	HGLYLPTR		0.8005	0.01287	0.5122	0.9961	1663546	
8	÷	2	HGLYLPTR	Γ	0.8910	0.04431	0.4473	0.9946	343135	
9	+	2	GLVQALQTK		0.8556	0.01614	0.9430	0.9976	1585877	
10	+	2	GLVQALQTK	Г	1.4824	0.06835	0.4623	0.9925	25356	
11	÷	2	TGTVSLEVR	Г	0.8174	0.02263	0.3872	0.9970	232042	
12	+	2	TGTVSLEVR	Г	1.1870	0.05471	0.2344	0.9793	27681	
13	+	2	VLEALLPLK		0.9445	0.02065	0.8568	0.9962	432986	
14	÷	2	VLEALLPLK		0.8698	0.00885	0.9534	0.9986	2234177	
15	+	2	VLEALLPLK		1.4595	0.04262	0.6748	0.9962	36608	
16	÷	2	VAAAVDLIIK		0.8341	0.01261	0.9118	0.9973	1506035	
17	÷	2	VAAAVDLIK	Γ	1.3130	0.04983	0.1966	0.9915	14575	
4.00		-	A CONTRACTOR DE LA CONTRA			0.0000	0.0000	0.0040		







Peak Lists Proteins Searches Acquisition Precursors





Differences in protein expression of human cells infected with Marburg viruses

1.12	mecian		
L/H	SD(geo)	#	Description
0.21	1.13	21	NAD(P)H dehydrogenase [quinone] 1 isoform a [Homo sapiens]
0.59	1.11	21	unnamed protein product [Homo sapiens]
0.59	1.16	10	ornithine aminotransferase, OAT
0.64	1.11	15	transketolase [Homo sapiens]
0.70	1.19	47	pyruvate carboxylase, mitochondrial precursor [Homo sapiens]
0.73	1.11	23	protein disulfide-isomerase A4 precursor [Homo sapiens]
0.77	1.21	10	microsomal triglyceride transfer protein [Homo sapiens]
0.78	1.17	10	estradioi 17-beta-dehydrogenase 12 [Homo sapiens]
0.79	1.08	12	chaperonin 10 [Homo sapiens]
0.79	1.08	77	60 kDa heat shock protein, mitochondrial [Homo sapiens]
1.59	1.33	10	drebrin E2 [Homo sapiens]
1.59	1.14	17	human rab GDI [Homo sapiens]
1.61	1.12	11	microtubule-associated protein RP/EB family member 1 [Homo se
1.63	1.11	11	cargo selection protein TIP47 [Homo sapiens]
1.63	1.11	18	microtubule-associated protein 1B [Homo sapiens]
1.63	1.09	23	vinculin isoform VCL [Homo sapiens]
1.65	1.09	18	microtubule-associated protein 4 isoform 1 [Homo sapiens]
1.69	1.09	14	peptidyi-prolyi cis-trans isomerase FKBP4 [Homo sapiens]
1.83	1.10	128	myosin-9 [Homo sapiens]
2.73	1.09	42	nestin, isoform CRA_c [Homo sapiens]

Pathway analysis of differential expression of proteins in human cells infected with Marburg viruses (SILAC data)



Bars indicate the total number of proteins (y-axis left) involved in the indicated pathway; green, upregulated in MARV-Ang ; red, upregulated in MARV-Mus. The line graph indicates the assigned –log(p-value) ratios (y-axis right) assessed via IPA for each respective pathway.

Differences in protein expression of human cells infected with Marburg viruses

1.12	Median		
L/H	SD(geo)	#	Description
0.09	6.00	11	VP30 [Lake Victoria marburgvirus - Angola2005]
0.39	328.10	20	RecName: Full=Matrix protein VP40
0.70	4.99	11	RecName: Full=Envelope glycoprotein
1.00	1.12	29	RecName: Full=Nucleoprotein
1.11	5.09	16	RecName: Full=Polymerase cofactor VP35
2.95	10.12	4	VP24 [Lake Victoria marburgvirus]
93.21	1.53	6	alpha-1B-glycoprotein precursor [Bos taurus]
97.45	7.10	131	PREDICTED: apolipoprotein B [Bos taurus]
100.50	9.57	4	PREDICTED: similar to complement component 4A [Bos taurus]
124.70	1.13	3	PREDICTED: pregnancy-zone protein-like [Bos taurus]
125.30	1.67	5	apolipoprotein A-I precursor [Bos taurus]
151.50	10.50	35	alpha-1-antiproteinase precursor [Bos taurus]
176.50	3.66	35	serotransferrin precursor [Bos taurus]
185.50	8.91	26	alpha-2-HS-glycoprotein precursor [Bos taurus]
253.20	4.06	6	transthyretin precursor [Bos taurus]
309.50	3.43	19	alpha-1-acid glycoprotein precursor [Bos taurus]

Differences in protein expression of human cells infected with lethal viruses using spectral counting and SILAC

	Total spectra				
Protein	Control	Ebola	Angola	Musoke	
fatty acid synthase	47	26	21	23	
beta-actin	29	32	42	42	
glyceraldehyde-3-phosphate dehydrogenase	13	14	19	20	

Significant differences were found for each protein by ANOVA (Scaffold)

Marburg-Angola (H), Marburg-Musoke (L)	SILAC L/H (SD geom.)					
Protein	Α	В	С			
Median	1.12	1.01	1.23			
fatty acid synthase	0.89 (1.41)	0.79 (1.51)	0.97 (1.16)			
beta-actin	1.27 (1.11)	1.14 (1.60)	1.40 (1.13)			
glyceraldehyde-3-phosphate dehydrogenase	1.06 (1.30)	0.91 (1.09)	1.13 (1.43)			

